Research Article

The essential oil composition of *Picea sitchensis* growing in the Oregon Coast Range

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Abstract

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Keywords

Sitka spruce, Pinaceae, gas chromatography, enantiomeric distribution, chiral *Picea sitchensis* (Sitka spruce) is a very large tree that is locally common in the moist coastal ranges of the Pacific Northwest. The purpose of this work was to obtain and analyze the foliar essential oil of the Sitka spruce growing in the Oregon Coast Range. Foliage from three individual trees was collected and the essential oils were obtained by hydrodistillation. The essential oils were analyzed by gas chromatographic techniques (GC-MS, GC-FID, and chiral GC-MS). The major components in the essential oils were α-pinene (1.2-12.5%, > 85% (–)-α-pinene), β-pinene (1.2-7.2%, > 95% (–)-β-pinene), myrcene (15.9-22.2%), β-phellandrene (9.1-25.1%, > 99% (–)-β-phellandrene), isoamyl isovalerate (2.3-6.4%), 3-methyl-3-butenyl isovalerate (1.5-6.9%), and piperitone (2.9-18.0%, > 98% (–)-piperitone). The essential oil compositions of the Oregon samples are qualitatively similar to samples from British Columbia. However, sampling from other populations at other latitudes and at different seasons of the year would be necessary to fully describe the volatile chemistry of this species.

1. Introduction

Picea sitchensis (Bong.) Carrière, Pinaceae (Sitka spruce) is a large tree (up to 80 m tall), with drooping branches (Fig. 1A) [1]. The leaves are needles (15-25 mm long), abaxial surface (blue-green), adaxial surface (glaucous) with two dense bands of stomata (Fig. 1B). The bark is thin, scaley, and grayish-brown in color (Fig. 1C). The cones are slender and cylindrical (6-10 cm long) with thin, diamond-shaped scales (12-16 mm) (Fig. 1D). Sitka spruce ranges along a narrow strip along the northern Pacific coast from south-central Alaska south to northern California (Fig. 2) [2, 3]. The tree was introduced to western Europe (Great Britain, France, Norway, Denmark, Germany, Sweden, and Iceland) in the 19th century [4, 5]. It has also been introduced to New Zealand and

Australia [6].

Picea species have been important sources of traditional medicines throughout their ranges (see, for example, [7–13]). The foliage of *P. sitchensis* has been used by Native American tribes as a cold medicine (Kwakiutl), as an antirheumatic (Gitksan), and as a gastrointestinal aid (Southern Carrier, Hanaksiala) [14]. Picea species, including Picea abies (L.) H. Karst (Norway spruce), Picea engelmannii Engelm. (Engelmann spruce), Picea glauca (Moench) Voss (white spruce), Picea mariana Britton, Sterns & Poggenb. (black spruce), Picea pungens Engelm. (blue spruce), Picea rubens Sarg. (red spruce), and P. sitchensis (Sitka spruce), are important sources of essential oils [15]. As part of our continuing





Figure 1. *Picea sitchensis* (Sitka spruce) in the Oregon Coast Range. A: Branches. B: Leaves (needles). C: Bark. D: Cones.



Figure 2. The native range of *Picea sitchensis* (Sitka spruce) along the west coast of North America [3].

investigation of essential oils of North American gymnosperms, we have obtained the foliar essential oils from three individual *P. sitchensis* trees growing in the Van Duzer Forest, Oregon Coast Range. There has been a previous report on *P. sitchensis* essential oils from coastal British Columbia [16], as well as a

report on volatiles identified in *P. sitchensis* bark extracts [17].

2. Materials and methods

2.1. Plant Material

Foliage (leaves and twigs) of P. sitchensis were collected using plant pruning shears from the ends of branches from several different positions on three individual trees (samples #1, #2, #3) located in the Van Duzer Forest, Oregon Coast Range on 14 April 2023. The trees were identified by W.N. Setzer based on botanical descriptions [18] and comparison with herbarium samples from the New York Botanical Garden [19]. A voucher specimen (WNS-Ps-6893) has been deposited in the University of Alabama in Huntsville herbarium. The fresh foliage from each tree was combined and the samples were stored under refrigeration (-20 °C). The fresh/frozen samples were hydrodistilled using a Likens-Nickerson apparatus with continuous extraction of the distillate with dichloromethane for four hours to give pale-yellow essential oils (Table 1).

2.2. Gas Chromatographic Analysis

The *P. sitchensis* foliar essential oils were analyzed by GC-MS, GC-FID, and chiral GC-MS as previously described [20]. The essential oil compositions were determined by comparing both MS fragmentation and RI values with those reported in the Adams [21], FFNSC3 [22], NIST20 [23], and Satyal [24] databases. The percent compositions were determined from raw peak areas (GC-FID) without standardization. Enantiomeric distributions were determined by comparison of RI values with authentic samples (Sigma-Aldrich, Milwaukee, WI, USA), which are compiled in our own in-house database.

3. Results and discussion

Hydrodistillation of the leaves (needles) and twigs of *P. sitchensis* gave pale-yellow essential oils in yields of 1.302-2.101%. The gas chromatographic analysis allowed for the identification of 140 chemical components, which accounted for 99.3-99.6% of the compositions (Table 2).

The foliar essential oil compositions of *P. sitchensis* from Oregon are qualitatively similar to those from British Columbia [16]. That is, the major components in both collections were α -pinene, β -pinene, myrcene, β -phellandrene, isoamyl isovalerate, 3-methyl-3-

Table 1. Collection and hydrodistillation details of Picea sitchensis from the Oregon Coast Range.

Samples	Location	Mass foliage	Mass essential oil	Yield (%)
#1	45°2'16" N, 123°48'29" W, 116 m asl	99.77 g	1.2731 g	1.302%
#2	45°2'16" N, 123°48'32" W, 116 m asl	116.25 g	2.4420 g	2.101%
#3	45°2'16" N, 123°48'34" W, 115 m asl	93.27 g	1.2648 g	1.356%

Table 2. Chemical compositions (%) of Picea sitchensis foliar	
essential oils from the Oregon Coast Range.	

		oni the Oregon Coast				RIcalc	RIdb	Compounds	#1	#2	#3
RIcalc	RIdb	Compounds	#1	#2	#3	1125	1124	cis-p-Menth-2-en-1-ol	1.3	0.4	0.8
780	780	(2Z)-Pentenol	0.1	0.2	0.2	1140	1141	trans-Pinocarveol	-	tr	-
700	792	3-Methyl-2-buten-1-	tr	0.5	0.2	1142	1142	trans-p-Menth-2-en-1-ol	1.0	0.3	0.6
762	782	ol (= Prenol)				1147	1145	Camphor	4.4	0.9	3.1
800	797	(3Z)-Hexenal	tr	0.1	0.2	1155	1156	Camphene hydrate	0.3	0.1	0.8
801	801	Hexanal	tr	0.2	0.3	1163	1162	iso-Borneol	-	-	tr
849	849	(2E)-Hexenal	0.9	2.1	2.9	1169	1169	Umbellulone	-	-	tr
850	853	(3Z)-Hexen-1-ol	0.1	0.4	0.4	1172	1170	Borneol	3.4	0.4	3.9
864	864	1-Hexanol	-	-	tr	1176	1176	cis-Pinocamphone	-	tr	0.1
872	873	Isoamyl acetate	-	-	tr	1150	1150	2-Isopropenyl-5-	tr	tr	0.1
880	880	Santene	-	0.1	tr	1178	1179	methyl-4-hexenal			
902	901	Heptanal	-	-	0.2	1180	1180	Terpinen-4-ol	0.4	0.4	0.8
922	923	Tricyclene	tr	tr	0.1	1100		Cyclopentyl 3-methyl-2-	0.1	tr	tr
925	925	α-Thujene	tr	0.1	0.3	1183	1184	butenoate			
932	933	α-Pinene	1.2	12.5	5.9	1187	1185	Cryptone	0.1	0.2	0.1
949	950	Camphene	0.6	0.6	1.8	1187	1186	<i>p</i> -Cymen-8-ol	0.1	tr	0.1
961	959	Benzaldehyde	0.1	tr	tr	1191	1192	Methyl salicylate	-	tr	-
967	967	Isoamyl propionate	tr	-	tr	1101	1100	2-Methyl-2-butenyl	0.2	-	-
972	971	Sabinene	0.2	0.3	0.7	1191	1190	angelate			
977	978	β-Pinene	1.2	7.2	4.4	1195	1195	α -Terpineol	1.0	0.9	1.6
989	989	, Mvrcene	22.2	15.9	18.2	1197	1196	<i>cis</i> -Piperitol	0.3	0.1	0.2
1007	1006	α-Phellandrene	0.5	0.4	0.5	1207	1208	Verbenone	-	0.1	_
1009	1008	δ-3-Carene	1.7	0.4	2.9	1209	1208	trans-Piperitol	0.6	0.1	0.3
1017	1017	α -Terpinene	0.2	0.2	0.3	1227	1227	Citronellol	0.1	_	tr
1024	1025	<i>p</i> -Cymene	0.6	0.4	0.4	1228	1229	Thymyl methyl ether	0.1	0.1	0.1
1029	1030	Limonene	0.8	1.1	0.7	1237	1238	Neral	0.1	-	tr
1031	1031	β-Phellandrene	9.1	25.1	17.5	1249	1252	Isoamyl hexanoate	0.2	0.1	0.1
1032	1032	1.8-Cineole	1.8	0.9	1.8	1250	1249	Geraniol	0.1	-	-
1034	1034	(Z)-β-Ocimene	tr	0.1	tr	1254	1254	Piperitone	18.0	29	5.8
1043	1043	Phenylacetaldehyde	tr	tr	tr	1258	1272	4-Pentenvl bevanoate ^a	0.1	0.1	0.1
1045	1045	(F) - β - Ω cimene	tr	tr	0.1	1268	12/2	Coranial	0.1	-	0.1 tr
1010	1010	Isoamul huturate	0.5	0.1	0.1	1200	1200	Phellandral	0.1	0.1	0.1
1051	1057	y-Terninene	0.2	0.1	0.5	1284	1277	Bornyl acetate	0.1	0.1	0.1
1007	1057	3-Methyl-2-butenyl	0.2	-	tr	1204	1202	2-Undecanone	-	-	tr
1063	1064	buturato	0.2	_	u	1292	1293	Mothyl myrtonato		0.1	u
		cie Sabinono	tr	0.1	0.1	1204	1204	(F) Cinnamyl alcohol	0.2	0.1	-
1070	1069	hydrate	u	0.1	0.1	1300	1304	4-Vinvlguaiacol	0.2	0.1	_
1085	1086	Torpipolopo	0.5	07	1.0	1302	1302	Mothyl docapoato	0.1		
1088	1000	Fenchone	0.5	0.7	0.1	1325	1322	cis-Piperitol acetate	0.1	0.1	0.1
1000	1090	67 Epoyumurcopo	0.1	0.1	0.1	1346	1346	a Cubabana	0.1	0.1	0.1 tr
1090	1090	Porillopo	0.1	0.1 tr	0.1 +r	1340	1265	(2E) Undecenal	-	0.1	11 +r
1090	1090	Linalool	0.1	0.2	0.2	1304	1275	(2L)-Ondecentar	- 0.1	0.2	0.2
1099	1101	tuqua Sabinana	0.4 +n	0.5	0.5	1375	1373	Coronal acotato	0.1 +n	0.2	0.2
1101	1101	hydrato	u	u	0.1	13/7	1370	B Cubabapa	u	11 ++	0.1
1104	1104	Noranal	L	0.2	0.2	1307	1307	p-Cubebene	-	0.1	u
1104	1104		tr 6.4	0.2	0.2	1434	1433	<i>cis</i> -mujopsene	-	U.1	-
1106	1109	(= Solution = 1)	0.4	2.3	3.2	1437	1439	2 Mather 2 but a 1	0.1	ır	0.1
		(= Solusterol)				1444	1443	3-MetnyI-2-buten-1-yl	-	-	0.1
1107	1109	2-Methylbutyl	-	tr	-			benzoate		0.1	
		isovalerate				1444	1446	<i>cis</i> -Muurola-3,5-diene	-	0.1	-
1116	1114	3-Methyl-3-butenyl	6.9	1.5	1.9	1448	1450	trans-Muurola-3,5-diene	-	0.1	tr
		isovalerate				1451	1452	(E)-β-Farnesene	-	0.1	tr
1120	1120	endo-Fenchol	tr	tr	0.8						

Table 2. (Continued)

RIcalc	RIdb	Compounds	#1	#2	#3
1461	1463	cis-Muurola-4(14),5-diene	tr	0.2	tr
1468	1467	9-epi-(E)-Caryophyllene	-	0.1	tr
1471	1472	Cadina-1(6),4-diene	-	0.1	tr
1474	1475	γ-Muurolene	-	0.2	0.1
1480	1480	Germacrene D	-	0.1	tr
1488	1487	β-Selinene	-	0.1	tr
1491	1490	γ-Amorphene	-	0.2	0.1
1495	1497	α -Selinene	0.1	0.4	0.1
1497	1500	α -Muurolene	tr	0.4	0.2
1502	1503	(E,E) - α -Farnesene	-	0.1	-
1511	1512	γ-Cadinene	0.1	1.2	0.4
1514	1515	Cubebol	-	0.1	0.1
1517	1518	δ-Cadinene	0.3	2.4	0.9
1521	1521	Zonarene	-	0.1	-
1531	1533	trans-Cadina-1,4-diene	-	0.1	tr
1535	1538	α -Cadinene	-	0.1	tr
1539	1540	(E)- α -Bisabolene	-	0.1	0.1
1559	1560	(E)-Nerolidol	0.1	-	-
1575	1576	Germacra-1(10),5-dien- 4β-ol	-	0.1	0.1
1602	1600	α -Oplopenone	-	0.2	0.1
1602	1593	Isoamyl 3-phenylpropio- nate	0.3	-	-
1608	1610	Cedrol	-	0.1	-
1613	1614	1,10-di-epi-Cubenol	0.1	0.1	tr
1627	1628	1-epi-Cubenol	-	0.1	0.1
1641	1640	τ-Cadinol	0.1	0.3	0.1
1643	1641	τ-Muurolol	0.1	0.3	0.1
1646	1646	Himachal-2-en-7β-ol	0.1	-	-
1654	1655	α-Cadinol	0.2	0.7	0.2
1662	1664	ar-Turmerone	tr	0.1	tr
1667	1668	α -Turmerone	0.1	0.4	0.1
1871	1875	Oplopanonyl acetate	1.7	0.5	0.2
1922	1929	Cembrene	0.5	1.3	1.6
1935	1931	Beyerene	0.6	0.5	0.3

butenyl isovalerate, and piperitone. Furthermore, the major monoterpenoids, α -pinene, β -pinene, δ -3carene, myrcene, and β-phellandrene, were also observed to be major components in the bark extracts from Vancouver Island, British Columbia [17]. A comparison of the main foliar essential oil components is summarized in Table 3. α -Pinene is a relatively abundant constituent in the foliar essential oils of *Picea* species [25–27]. On the other hand, β pinene, myrcene, β-phellandrene, isoamyl isovalerate, 3-methyl-3-butenyl isovalerate, and piperitone were not present in one or more Picea species [25, 28, 29]. Bornyl acetate is often an abundant constituent of *Picea* essential oils [25, 26], but was not observed in P. engelmannii from northern Arizona [28]. Similarly, camphor is often found in Picea essential oils, but was not detected in samples of P. glauca [25] or P. sitchensis [16] from Canada.

Table 2. (Continued)									
RIcalc	RIdb	Compounds	#1	#2	#3				
1936	1934	(3Z)-Cembrene A	0.1	0.4	0.6				
1946	1946	<i>m</i> -Camphorene	0.1	-	tr				
1953	1951	(3E)-Cembrene A	-	0.1	0.1				
1965	1968	Sandaracopimara-	-	0.1	Tr				
1705	1700	8(14),15-diene							
1993	1994	Manoyl oxide	0.1	0.1	0.1				
1997	2000	Isopimara-7,15-diene	-	0.2	0.1				
2004	1998	Luxuriadiene	0.2	0.1	-				
2041	2038	Thunbergol A	0.3	0.5	0.9				
2053	2053	Manool	4.3	1.8	2.7				
2085	2086	Abietadiene	0.2	0.5	0.4				
2222	2224 ^b	Isopimarinal	-	-	0.1				
2230	2245°	Palustral	0.5	1.4	0.9				
2234	2265 ^c	Levopimarinal	0.1	0.3	0.1				
2262	2266	Dehydroabietal	0.1	0.1	0.1				
2307	2312	Abietal	0.1	0.3	0.1				
2366	2366	Neoabietic acid	0.1	0.2	0.1				
Compo	ound Cla	sses							
Monot	erpene h	ydrocarbons	38.8	65.4	55.3				
Oxyge	nated m	33.9	8.4	22.6					
Sesquiterpene hydrocarbons				6.4	2.0				
Oxyge	nated se	2.3	3.1	1.0					
Diterp	enoids	7.2	8.0	8.2					
Benzei	noid aror	0.4	0.1	0.2					
Others	6	16.0	8.1	10.2					
Total	identified	99.3	99.4	99.6					

 RI_{calc} = Retention index calculated with respect to a homologous series of *n*-alkanes on a ZB-5ms column. RI_{db} = Retention index from the available databases [21–24] unless otherwise indicated. tr = trace (< 0.05%). ^a The identification is only tentative; although there is a good MS match, the RI values are very different. ^b RI value from Shpatov et al., 2017 [38]. ^c The identification is only tentative; although there is a good MS match, the RI values are very different; however, the compound was identified in the bark of *P. sitchensis* [17].

Although the essential oils are qualitatively similar, there are some notable quantitative differences. Monoterpene hydrocarbons were generally higher in the British Columbia samples compared to those from Oregon, while oxygenated monoterpenoid and sesquiterpenoid concentrations were higher in the Oregon samples. It is not clear what factors affect the compositional differences. Previous workers have reported large variations in monoterpene concentrations in essential oils of Picea species both within and between populations [27, 30] as well as seasonal variations in individual monoterpenoid concentrations Furthermore, [31-33]. the monoterpenoid concentrations vary widely between young leaves and older leaves in P. sitchensis; myrcene was found in high concentrations in immature foliage (95%), but decreased with age with concomitant increase in piperitone concentration [34]. The volatile

Table 3. Comparison of the percentages of the main components in the essential oils of *Picea sitchensis* from Oregon, British Columbia, and a commercial sample from New Zealand.

Compounds	Oregon (f	this work)	British Co	British Columbia [16]		
	Average	Range	Average	Range	Commercial ^a	
α-Pinene	6.5	1.2-12.5	7.4	2.9-11.5	5.1	
Camphene	1.0	0.6-1.8	1.0	0.0-1.6	3.0	
β-Pinene	4.3	1.2-7.2	5.6	2.9-9.8	3.3	
Myrcene	18.8	15.9-22.2	23.1	12.1-33.3	20.7	
δ-3-Carene	1.7	0.4-2.9	2.9	0.0-5.9	2.0	
Limonene	0.8	0.7-1.1	4.5	1.3-9.9	10.8	
β-Phellandrene	17.2	9.1-25.1	21.1	15.5-35.6	9.9	
1,8-Cineole	1.5	0.9-1.8	1.3	1.2-1.8	1.7	
Terpinolene	0.7	0.5-1.0	0.9	0.0-1.7	0.9	
Isoamyl isovalerate (= Solusterol)	4.0	2.3-6.4	3.4	0.8-6.4	3.8	
3-Methyl-3-butenyl isovalerate	3.5	1.5-6.9	2.3	0.0-5.3	1.4	
Camphor	2.8	0.9-4.4	2.2	0.0-3.5	23.8	
Borneol	2.6	0.4-3.9	0.7	0.0-1.8	2.2	
α -Terpineol	1.2	0.9-1.6	0.6	0.0-1.0	0.7	
Piperitone	8.9	2.9-18.0	7.0	0.5-12.5	2.2	
δ-Cadinene	1.2	0.3-2.4	1.3	0.3-4.2	0.1	
Manool	3.0	1.8-4.3	1.7	0.3-3.3	0.2	
Monoterpene hydrocarbons	53.2	38.8-65.4	77.1	68.5-90.4	58.2	
Oxygenated monoterpenoids	21.6	8.4-33.9	9.9	4.5-14.6	35.1	
Sesquiterpenoids	5.2	2.9-9.5	2.8	1.3-3.6	0.3	

^a Unpublished data from the Aromatic Plant Research Center, Lehi, Utah, USA.

components of Picea species play an important role in avoidance of insects and browsing by herbivores. In P. sitchensis, both myrcene and piperitone affect the feeding behavior of spruce aphids (Cinara costata, Cinara pilicornis, Cinara pruinosa, and Elatobium abietinum) depending on their tolerance to myrcene or piperitone [30]. Total monoterpene concentration was shown to negatively influence browsing of P. sitchensis by red deer (Cervus elaphus) [35]. Genetic, edaphic, climatic, and geographic factors are often cited as affecting the essential oil profiles [36, 37]. Latitudinal differences, including climatic differences, may be responsible for the lower monoterpene hydrocarbon concentrations and higher oxygenated monoterpenoid concentrations in the Oregon samples compared to the British Columbia samples. A commercial sample of P. sitchensis essential oil from New Zealand (unpublished data from the Aromatic Plant Research Center, Lehi, Utah, USA) has also been included in Table 3 for comparison. The concentration of limonene in the New Zealand sample was notably higher, with concomitant lower β -phellandrene, than in the North American samples. The concentration of camphor was also very high in the New Zealand sample, while the total sesquiterpenoids was very low.

The Oregon P. sitchensis foliar essential oils were also analyzed by chiral GC-MS in order to determine the distributions enantiomeric of the terpenoid components (Table 4). The levorotatory enantiomers were dominant for α -pinene, β -pinene, limonene, β phellandrene, borneol, and piperitone, while (+)camphor, $(+)-\delta$ -3-carene, and $(+)-\delta$ -cadinene were dominant. Camphene, linalool, terpinen-4-ol, and α terpineol were virtually racemic. Robert [17] also found (–)- α -pinene, (–)- β -pinene, (+)- δ -3-carene, and (-)- β -phellandrene to dominate the bark extract of *P*. sitchensis. However, (+)-limonene rather than (-)limonene was identified in the bark extract. Consistent with the distributions in *P. sitchensis*, (–)- α pinene, (-)- β -pinene, (-)-limonene, and $(-)-\beta$ phellandrene were the predominant enantiomers in P. pungens [26] and P. engelmannii [27].

4. Conclusions

This work presents the first report on the foliar essential oil of *Picea sitchensis* from the Oregon Coast Range and includes the enantiomeric distributions of chiral terpenoid components. The essential oils of *P. sichensis* were rich in (-)- α -pinene, (-)- β -pinene, myrcene, (-)- β -phellandrene, isoamyl isovalerate, 3-methyl-3-butenyl isovalerate, and (-)-piperitone.

Table 4. Enantiomeric distribution (%) of terpenoidcomponents in *Picea sitchensis* from the Oregon Coast Range.

Compounds	RIdb	RIcalc	#1	#2	#3
(+)-α-Thujene	950	-	-	-	0.0
(–)-α-Thujene	951	951	-	-	100.0
(–)-α-Pinene	976	975	86.1	95.2	92.0
(+)-α-Pinene	982	982	13.9	4.8	8.0
(–)-Camphene	998	1000	54.7	60.0	66.8
(+)-Camphene	1005	1004	45.3	40.0	33.2
(+)-Sabinene	1021	1021	-	-	14.2
(–)-Sabinene	1030	1029	-	-	85.8
(+)-β-Pinene	1027	1027	5.0	3.3	3.1
(–)-β-Pinene	1031	1031	95.0	96.7	96.9
(+)-δ-3-Carene	1052	1052	100.0	100.0	100.0
(–)-δ-3-Carene	na	-	0.0	0.0	0.0
(–)-Limonene	1073	1079	77.8	76.7	70.2
(+)-Limonene	1081	1082	22.2	23.3	29.8
(–)-β-Phellandrene	1083	1083	99.7	99.6	99.6
(+)-β-Phellandrene	1089	1087	0.3	0.4	0.4
(–)-Linalool	1228	1216	43.5	48.9	60.0
(+)-Linalool	1231	1220	56.5	51.1	40.0
(–)-Camphor	1253	1253	2.1	0.0	4.6
(+)-Camphor	1259	1255	97.9	100.0	95.4
(+)-Terpinen-4-ol	1297	1298	58.3	56.6	42.0
(–)-Terpinen-4-ol	1300	1301	41.5	43.4	58.0
(–)-Borneol	1335	1338	70.0	56.9	74.5
(+)-Borneol	1340	1348	30.0	43.1	25.5
(–)-α-Terpineol	1347	1350	36.7	55.5	40.4
(+)- α -Terpineol	1356	1358	63.3	44.5	59.6
(-)-Piperitone	1380	1385	99.2	98.3	99.1
(+)-Piperitone	1385	1391	0.8	1.7	0.9
(–)-δ-Cadinene	1563	-	0.0	0.0	0.0
(+)-δ-Cadinene	1576	1567	100.0	100.0	100.0

RI_{db} = Retention index from our in-house database developed using commercially available samples on a Restek B-Dex 325 column. RI_{calc} = Retention index determined with respect to a homologous series of *n*-alkanes on a Restek B-Dex 325 column. na = reference compound not available. - = compound not observed.

While the compositions are qualitatively similar to those from British Columbia, it would be interesting to compare the Oregon and British Columbia essential oil profiles with populations from Washington state and northern California as well as western Europe in order to more fully appreciate the quantitative differences based on geographical location. Since *P. sichensis* is commercially important and cultivated for lumber, the foliage recovered represents a value-added commodity, which may be commercially exploited in the essential oil industry.

Authors' contributions

Conceptualization, W.N.S.; Methodology, P.S. and

W.N.S.; Software, P.S.; Validation, W.N.S., Formal Analysis, P.S., A.P., and W.N.S.; Investigation, P.S., A.P., K.S., and W.N.S.; Resources, P.S. and W.N.S.; Data Curation, W.N.S.; Writing – Original Draft Preparation, W.N.S.; Writing – Review & Editing, P.S., A.P., K.S., and W.N.S.; Project Administration, W.N.S.

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Availability of data and materials

All data will be made available on request according to the journal policy.

Conflicts of interest

The authors declare no conflict of interest.

References

- Flora of North America Editorial Committee, F.N.A.E.C. *Picea sitchensis* (Bongard) Carrière Available online: http://www.efloras.org/florataxon.aspx?flora_id=1&ta xon_id=233500914 (accessed on Nov 11, 2023).
- Harris, A.S. *Picea sitchensis* (Bong.) Carr. Sitka Spruce. In Silvics of North America, Volume 1. Conifers; Burns, R.M., Honkala, B.H., Eds.; Forest Service, United States Department of Agriculture: Washington, DC, USA, 1990; pp. 260–267.
- Little, E.L. Digital representations of tree species range maps Available online: https://commons.wikimedia.org/w/index.php?curid=2 9262734 (accessed on Nov 11, 2023).
- Mason, B.; Perks, M.P. Sitka spruce (*Picea sitchensis*) forests in Atlantic Europe: Changes in forest management and possible consequences for carbon sequestration. Scand. J. For. Res. 2011, 26, 72–81, https://doi.org/10.1080/02827581.2011.564383.
- Nygaard, P.H.; Øyen, B.H. Spread of the introduced Sitka spruce (*Picea sitchensis*) in coastal Norway. Forests 2017, 8, 24, https://doi.org/10.3390/f8010024.
- 6. O-Driscoll, J. Sitka spruce, its distribution and genetic variation. Irish For. J. 1977, 34, 1–16.
- 7. Turner, N.J. Ethnobotany of coniferous trees in Thompson and Lillooet Interior Salish of British

Columbia. Econ. Bot. 1988, 42, 177–194.

- Kirn, H.S.; Kapahi, B.K.; Srivastava, T.N. Taxoethnobotanical observations on the gymnosperms of Poonch district (J. &. K state) India. J. Econ. Taxon. Bot. 1999, 23, 155–160.
- Leduc, C.; Coonishish, J.; Haddad, P.; Cuerrier, A. Plants used by the Cree Nation of Eeyou Istchee (Quebec, Canada) for the treatment of diabetes: A novel approach in quantitative ethnobotany. J. Ethnopharmacol. 2006, 105, 55–63, https://doi.org/10.1016/j.jep.2005.09.038.
- Šarić-Kundalić, B.; Dobeš, C.; Klatte-Asselmeyer, V.; Saukel, J. Ethnobotanical study on medicinal use of wild and cultivated plants in middle, south and west Bosnia and Herzegovina. J. Ethnopharmacol. 2010, 131, 33–55, https://doi.org/10.1016/j.jep.2010.05.061.
- Kayani, S.; Ahmad, M.; Sultana, S.; Khan Shinwari, Z.; Zafar, M.; Yaseen, G.; Hussain, M.; Bibi, T. Ethnobotany of medicinal plants among the communities of Alpine and Sub-alpine regions of Pakistan. J. Ethnopharmacol. 2015, 164, 186–202, https://doi.org/10.1016/ j.jep.2015. 02.004.
- Kazancı, C.; Oruç, S.; Mosulishvili, M. Medicinal ethnobotany of wild plants: a cross-cultural comparison around Georgia- Turkey border, the Western Lesser Caucasus. J. Ethnobiol. Ethnomed. 2020, 16, 71, https://doi.org/10.1186/s13002-020-00415-y.
- Thapa-Magar, D.K.; Thapa-Magar, K.B.; Kunwar, R.M.; Bussmann, R.W.; Paniagua-Zambrana, N.Y.; Hussain, W. *Picea smithiana* (Wall.) Boiss. Pinaceae. In Ethnobotany of the Himalayas; Kunwar, R.M., Sher, H., Bussmann, R.W., Eds.; Springer International Publishing: Cham, Switzerland, 2021; pp. 1507–1514 ISBN 9783030455972.
- Moerman, D.E. Native American Ethnobotany; Timber Press, Inc.: Portland, Oregon, USA, 1998; ISBN 978-0-88192-453-4.
- 15. Rhind, J.P. Essential Oils: A Comprehensive Handbook for Aromatic Therapy; Singing Dragon: London, UK, 2020; ISBN 978-1787752290.
- St-Gelais, A.; Collin, G.; Helbig, J.; Gagnon, H. Essential oils from the foliage of *Picea sitchensis* from British Columbia. Am. J. Essent. Oils Nat. Prod. 2018, 6, 19–26.
- 17. Robert, J.A. Terpenoid Profiling and Biosynthesis in Sitka Spruce (*Picea sitchensis*) Genotypes That Are Susceptible or Resistant to Attack by the White Pine Weevil (*Pissodes strobi*), Ph.D. thesis, University of British Columbia, 2010.
- Turner, M.; Kuhlmann, E. Trees & Shrubs of the Pacific Northwest; Timber Press, Inc.: Portland, Oregon, USA, 2014; ISBN 978-1-60469-263-1.
- 19. New York Botanical Garden, N.Y.B.G. C. V. StarrVirtualHerbariumAvailableonline:

https://sweetgum.nybg.org/science/vh/specimenlist/?SummaryData=Picea sitchensis (accessed on Apr 18, 2023).

- Satyal, P.; Dosoky, N.S.; Poudel, A.; Swor, K.; Setzer, W.N. Chemical composition of the aerial parts essential oil of *Chrysothamnus viscidiflorus* from southwestern Idaho. J. Essent. Oil Plant Compos. **2023**, *1*, 115–121, https://doi.org/10.58985/jeopc.2023.v01i02.16.
- 21. Adams, R.P. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry; 4th ed.; Allured Publishing: Carol Stream, Illinois, USA, 2007; ISBN 978-1-932633-21-4.
- 22. Mondello, L. FFNSC 3; Shimadzu Scientific Instruments: Columbia, Maryland, USA, 2016.
- 23. NIST20; National Institute of Standards and Technology: Gaithersburg, Maryland, USA, 2020.
- 24. Satyal, P. Development of GC-MS Database of Essential Oil Components by the Analysis of Natural Essential Oils and Synthetic Compounds and Discovery of Biologically Active Novel Chemotypes in Essential Oils, Ph.D. dissertation, University of Alabama in Huntsville, Huntsville, AL, USA, 2015.
- Koçak, A.; Kılıç, Ö. Identification of essential oil composition of four *Picea* Mill. (Pinaceae) species from Canada. J. Agric. Sci. Technol. B 2014, 4, 209–214.
- Swor, K.; Poudel, A.; Satyal, P.; Setzer, W.N. Chemical composition and enantiomeric distribution of *Picea pungens* essential oil. Am. J. Essent. Oils Nat. Prod. 2021, 9, 14–18.
- 27. Swor, K.; Satyal, P.; Poudel, A.; Setzer, W.N. Gymnosperms of Idaho: Chemical compositions and enantiomeric distributions of essential oils of *Abies lasiocarpa*, *Picea engelmannii*, *Pinus contorta*, *Pseudotsuga menziesii*, and *Thuja plicata*. Molecules 2023, 28, 2477, https://doi.org/10.3390/molecules28062477.
- Wagner, M.R.; Clancy, K.M.; Tinus, R.W. Maturational variation in needle essential oils from *Pseudotsuga* menziesii, Abies concolor and Picea engelmannii. Phytochem. 1989, 28, 765–770, https://doi.org/10.1016/ 0031-9422(89)80111-6.
- Mardarowicz, M.; Wianowska, D.; Dawidowicz, A.L.; Sawicki, R. Comparison of terpene composition in Engelmann spruce (*Picea engelmannii*) using hydrodistillation, SPME and PLE. Zeitschrift fur Naturforsch. Sect. C J. Biosci. 2004, 59, 641–648, https://doi.org/10.1515/znc-2004-9-1006.
- Jackson, D.L.; Jarosik, V.; Dixon, A.F.G. Resource partitioning and tolerance of monoterpenes in four species of spruce aphid. Physiol. Entomol. 1996, 21, 242–246, https://doi.org/10.1111/j.1365-3032.1996.tb008 61.x.
- 31. Baath, H.M.; Burzo, I. Quantitative and qualitative seasonal variation of volatile oil from 16 conifer species.

Analele științifice ale Univ. "Al. I. Cuza" Iași 2009, 2, 103–110.

- 32. Kamaitytė-Bukelskienė, L.; Ložienė, K.; Labokas, J. Dynamics of isomeric and enantiomeric fractions of pinene in essential oil of *Picea abies* annual needles during growing season. Molecules. 2021, 26, 2138, https://doi.org/10.3390/molecules26082138.
- 33. Schoss, K.; Kočevar Glavač, N.; Kreft, S. Volatile compounds in Norway spruce (*Picea abies*) significantly vary with season. Plants. 2023, 12, 188, https://doi.org/10.3390/plants12010188.
- Hrutfiord, B.F.; Hopley, S.M.; Gara, R.I. Monoterpenes in Sitka spruce: Within tree and seasonal variation. Phytochem. 1974, 13, 2167–2170, https://doi.org/10. 1016/0031-9422(74)85021-1.
- 35. Duncan, A.J.; Hartley, S.E.; Thurlow, M.; Young, S.; Staines, B.W. Clonal variation in monoterpene concentrations in Sitka spruce (*Picea sitchensis*) saplings and its effect on their susceptibility to browsing damage by red deer (*Cervus elaphus*). For. Ecol. Manage. 2001, 148, 259–269, https://doi.org/10.1016/S0378-1127(00)00540-5.

- 36. Karimi, A.; Krähmer, A.; Herwig, N.; Schulz, H.; Hadian, J.; Meiners, T. Variation of secondary metabolite profile of *Zataria multiflora* Boiss. populations linked to geographic, climatic, and edaphic factors. Front. Plant Sci. 2020, 11, 969, https://doi.org/10.3389/fpls.2020.00969.
- Vaičiulytė, V.; Ložienė, K.; Taraškevičius, R. Impact of edaphic and climatic factors on *Thymus pulegioides* essential oil composition and potential prevalence of chemotypes. Plants. 2022, 11, 2536, https://doi.org/10.3390/plants11192536.
- Shpatov, A. V.; Popov, S.A.; Salnikova, O.I.; Kukina, T.P.; Shmidt, E.N.; Um, B.H. Composition and bioactivity of lipophilic metabolites from needles and twigs of Korean and Siberian pines (*Pinus koraiensis* Siebold & Zucc. and *Pinus sibirica* Du Tour). Chem. Biodivers. 2017, 14, e1600203. https://doi.org/10.1002/ cbdv.201600203.