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Volatile components of the aerial parts of *Prunella vulgaris* L. (Lamiaceae)

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Abstract

Prunella vulgaris (“heal-all”) is found throughout the northern hemisphere and has been used as a traditional medicine in many cultures. In this work, the aerial parts of *P. vulgaris*, growing wild in north Alabama, were collected and hydro distilled to give the essential oil, which was analyzed by gas chromatography – mass spectrometry. The major components in the essential oil were germacrene D (49.7%), β-barbatene (9.0%), hexanal (5.1%), (3Z)-hexenol (4.6%), and cuparene (4.5%). The essential oil of *P. vulgaris* was screened for antifungal activity against *Aspergillus niger*, *Candida albicans*, and *Cryptococcus neoformans*, but showed negligible activity (MIC = 625, 1250, and 156 µg/mL, respectively).

Keywords: Essential oil composition, germacrene, barbatene

1. Introduction

Prunella vulgaris L., “heal-all” or “self-heal” (Lamiaceae), is a perennial Holarctic herb and can be found throughout North America [1, 2]. The plant serves as a traditional herbal medicine in several cultures. For example, it has been used in Chinese traditional medicine for “heat clearing” effects [3, 4], in Kashmir, the plant is used in Unani medicine as a “brain tonic” and to treat symptoms of colds [5], and in Europe, the herb has been used externally to treat wounds and internally as a tonic and as a gargle for sore throat and mouth sores [1, 6]. Native Americans have also used *P. vulgaris* as a traditional medicine. The Blackfoot used an infusion of the plant on sores, the Cherokee used an infusion on burns, and the Iroquois took an infusion internally as a tonic and to treat backache [7].

Phytochemical studies of *P. vulgaris* have shown the plant to be rich in triterpenoids and phenolics [8]. As part of our ongoing interest in essential oils from Cherokee aromatic medicinal plants [8-14], we have examined the essential oil composition from the aerial parts of *P. vulgaris* collected in north Alabama.

2. Materials and methods

2.1 Plant material

The aerial parts of *P. vulgaris* were collected on 15 September 2018 near the community of Gurley in north Alabama (34°39'19.2"N, 86°24'47.4"W, elev. 215 m). The plant was identified by S.K. Lawson; a voucher specimen (20190401-000952) has been deposited in the University of Alabama in Huntsville herbarium. The fresh plant material (22.47 g) was hydrodistilled using a Likens-Nickerson apparatus, with continuous extraction with CH₂Cl₂, for 3 h to give a pale-yellow essential oil (5.0 mg).

2.2 Gas Chromatographic – mass spectral analysis

The essential oil of *P. vulgaris* was analyzed by GC-MS, as described previously [15, 16], using a Shimadzu GC-MS-QP2010 Ultra fitted with a Phenomenex ZB-5ms column. Identification of the essential oil components was determined by comparison of their retention indices and their mass spectral fragmentation patterns with those in the essential oil databases [17-20].

2.3 Antifungal screening

The *C. coelestinum* essential oil was screened for antifungal activity against *Aspergillus niger* (ATCC 16888), *Candida albicans* (ATCC 18804), and *Cryptococcus neoformans* (ATCC 24607) using the micro broth dilution method as previously described [21]. Amphotericin B was used as the positive control and RPMI medium was used as the negative control.

3. Results and discussion

The aerial parts of *P. vulgaris* were collected from wild-growing plants in north Alabama and

hydrodistilled to give a pale-yellow essential oil in 0.022% yield. Twenty-seven compounds representing 99.2% of the composition were identified in the essential oil (see Table 1).

The sesquiterpene germacrene D (49.7%) dominated the essential oil with lesser concentrations of β -barbatene (9.0%), hexanal (5.1%), (3Z)-hexenol (4.6%), and cuparene (4.5%).

Table 1: Aerial parts essential oil composition of *Prunella vulgaris* L

RI ^a	Compound	%	RI ^a	Compound	%
795	1-Methylhept-2-ene	0.4	1480	Germacrene D	49.7
800	Octane	0.8	1494	Bicyclogermacrene	2.5
801	Hexanal	5.1	1505	α -Chamigrene	0.6
810	2-Hexanol	0.5	1507	Cuparene	4.5
850	(2E)-Hexenal	1.9	1517	δ -Cadinene	0.8
851	(3Z)-Hexenol	4.6	1533	γ -Cuprenene	1.5
864	1-Hexanol	1.5	1576	Spathulenol	2.4
932	α -Pinene	0.5	1654	α -Cadinol	0.6
1028	Limonene	1.0	1829	Unidentified sesquiterpenoid ^b	0.8
1043	Benzene acetaldehyde	1.4	1839	Phytone	1.4
1383	β -Bourbonene	1.3		Monoterpene hydrocarbons	1.5
1388	β -Elemene	2.9		Sesquiterpene hydrocarbons	76.6
1411	2,5-Dimethoxy- <i>p</i> -cymene	0.5		Oxygenated sesquiterpenoids	3.0
1414	α -Barbatene	1.8		Diterpenoids	1.4
1417	β -Ylangene	0.9		Benzenoids	1.9
1429	β -Copaene	0.7		Others	14.8
1434	<i>cis</i> -Thujopsene	0.5		Total Identified	99.2
1448	β -Barbatene	9.0			

^a Retention Index determined with reference to a homologous series of *n*-alkanes on a ZB-5ms column

^b207(6%), 191(100%), 163(40%), 151(71%), 146(37%), 131(43%), 123(74%), 107(67%), 105(61%), 95(60%), 93(77%), 91(71%), 83(95%), 81(76%), 79(57%), 77(54%), 69(61%), 67(53%), 55(52%), 43(28%), 41(47%)

The aerial parts essential oil of *P. vulgaris* from north Alabama showed a chemical composition markedly different from previous reports from Iran [22] and from China [23-24].

Chen and co-workers reported that the leaf essential oil of *P. vulgaris* collected from Anhui province, China, was composed mainly of aromadendrene (55.4%), cucumber alcohol (8.5%) and phytol (5.1%) [23]. This analysis is doubtful, however. The Kovats retention indices were not reported, but the retention time of "aromadendrene" was between those of limonene and linalool on a DB-5 column, and there are several other compounds listed that are out of order for elution from a DB-5 column. Yang and co-workers used headspace solid-phase micro extraction (HS-SPME) followed by GC-MS to analyze *P. vulgaris* aerial parts from Changchun, China, and found phytone (12.5%), α -santalene (11.2%), α -bourbonene (5.7%), geranyl acetone (5.7%), and caryophyllene oxide (5.1%) to be the major components [24]. The identification of geranyl acetone and α -santalene are in doubt, however; the reported RI values (1446 and 1453, respectively on a DB-5 column) are too different from those in the Adams database (1453 and 1416, respectively on a DB-5 column) [17].

The essential oil of *P. vulgaris* was screened for antifungal activity against *Aspergillus niger*, *Candida albicans*, and *Cryptococcus neoformans*, but showed negligible activity (MIC = 625, 1250, and 156 μ g/mL, respectively).

4. Conclusions

The essential oil compositions from *P. vulgaris* from different geographical areas vary widely. The chemical differences are likely to have profound effects on the biological activities of this herbal medicine and should be considered. Since the essential oil compositions of *P. vulgaris* reported in the literature are in doubt, it would be useful for additional analysis of this herbal medicinal plant from other geographical areas to be carried out.

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6. Conflicts of Interest

The authors declare no conflicts of interest.

7. References

- Foster S, Duke, J.A. A Field Guide to Medicinal Plants. Houghton Mifflin, Boston, Massachusetts, USA, 1990.
- Kartesz JT. The biota of North America program (BONAP). Available online: <http://bonap.net/MapGallery/County/Prunellavulgaris.png> (accessed on Aug 13, 2019).
- Du D, Cheng Z, Chen D. A new unusual $\Delta^{11(12)}$ -Oleane triterpene and anti-complementary triterpenes from *Prunella vulgaris* spikes natural product communications. 2012; 7(4):501-505.
- Gu XJ, Li YB, Li P, Qian SH, Zhang JF. Triterpenoids saponins from the spikes of *Prunella vulgaris*, Helvetica Chimica Acta. 2007; 90:72-78.
- Rasool R, Ganai BA, Akbar S, Kamili AN, Masood A. Phytochemical screening of *Prunella vulgaris* L. An important medicinal plant of Kashmir, Pakistan Journal of pharmaceutical sciences. 2010; 23(4):399-402.
- Grieve M. A Modern Herbal, Dover Publications, New York, NY, USA, 1971, 2.
- Moerman DE. Native American Ethno botany. Timber Press, Portland, OR, USA, 1998.
- Setzer W. The Phytochemistry of Cherokee aromatic medicinal plants medicines. 2018; 5(4):121.
- Satyral P, Craft JD, Dosoky NS, Setzer WN. The chemical compositions of the volatile oils of garlic (*Allium sativum*) and wild garlic (*Allium vineale*), Foods. 2017; 6(8):63.
- Woods KE, Chhetri BK, Jones CD, Goel N, Setzer WN. Bioactivities and compositions of *Betula nigra* essential oils, Journal of medicinally active plants. 2013; 2(1):1-9.
- Steinberg KM, Satyal P, Setzer WN. Bark essential oils

- of *Zanthoxylum clava-herculis* and *Ptelea trifoliata*: Enantiomeric distribution of meroterpenoids, Natural product communications. 2017; 12(6):961–963.
12. Stewart CD, Jones CD, Setzer WN. Essential oil compositions of *Juniperus virginiana* and *Pinus virginiana*, two important trees in Cherokee traditional medicine, American Journal of essential oils and natural products. 2014; 2(2):17–24.
 13. Setzer WN. Chemical composition of the leaf essential oil of *Lindera benzoin* growing in North Alabama, American Journal of essential oils and natural products. 2016; 4(3):1–3.
 14. Stewart CD, Jones CD, Setzer WN. Leaf essential oil compositions of *Rudbeckia fulgida* Aiton, *Rudbeckia hirta* L, and *Symphytotrichum novae-angliae* (L.) G.L, Nesom (Asteraceae), American Journal of essential oils and natural products. 2014; 2(1):36–38.
 15. DeCarlo A, Johnson S, Okeke-Agulu KI, Dosoky NS, Wax SJ, Owolabi MS *et al.* Compositional analysis of the essential oil of *Boswellia dalzielii* frankincense from West Africa reveals two major chemo types Phytochemistry. 2019; 164:24-32.
 16. Vargas Suarez A, Satyal P, Setzer WN. The wood essential oil composition of *Swietenia macrophylla* from Guanacaste, Costa Rica, American Journal of essential oils and natural products. 2019; 7(1):14–16.
 17. Adams RP. Identification of essential oil components by gas chromatography/mass spectrometry, 4th ed., Allured Publishing, Carol Stream, IL, USA, 2007.
 18. NIST17. National Institute of Standards and Technology, Gaithersburg, Maryland, USA, 2017.
 19. Mondello L. FFNSC 3. Shimadzu Scientific Instruments, Columbia, Maryland, USA, 2016.
 20. Satyal P. Development of GC-MS Database of Essential Oil Components by the Analysis of Natural Essential Oils and Synthetic Compounds and Discovery of Biologically Active Novel Chemotypes in Essential Oils. Ph.D. Dissertation, University of Alabama in Huntsville, 2015.
 21. Powers CN, Osier JL, McFeeters RL, Brazell CB, Olsen EL, Moriarity DM *et al.* Antifungal and cytotoxic activities of sixty commercially-available essential oils molecules. 2018; 23(7):1549.
 22. Morteza-Semnani K, Saeedi M, Akbarzadeh M. The essential oil composition of *Prunella vulgaris* L, Journal of essential oil-bearing plants. 2006; 9(3):257-260.
 23. Chen Y, Guo Q, Zhu Z, Zhang L, Dai X. Comparative analysis of the essential oil of flowers, leaves and stems of *Prunella vulgaris* L, Journal of essential oil-bearing plants. 2012; 15(4):662–666.
 24. Yang Y, Nan H, Wang G, Yang W, Xu J. Comparative determination of the volatile components of *Prunella vulgaris* L from different geographical origins by headspace solid-phase micro extraction and gas chromatography-mass spectrometry analytical letters. 2013; 46(13):2001-2016.