

American Journal of Essential Oils and Natural Products

Available online at www.essencejournal.com

American Journal of Essential Oils and Natural Products

J

E

0

N

р

ISSN: 2321 9114 AJEONP 2016; 4(3): 01-03 © 2016 AkiNik Publications Received: 01-05-2016 Accepted: 02-06-2016

William N Setzer

Department of Chemistry, University of Alabama in Huntsville Huntsville, AL 35899, USA

Chemical composition of the leaf essential oil of *Lindera* benzoin growing in North Alabama

William N Setzer

Abstract

The leaf essential oil of *Lindera benzoin* (L.) Blume (Lauraceae), growing in Huntsville, Alabama, was isolated by hydrodistillation. This plant, known commonly as spicebush, was used in native American traditional medicine, and is a host plant of the spicebush swallowtail butterfly, *Papilio troilus*. The chemical composition of the leaf oil of *L. benzoin* was determined by GC–MS. The most abundant essential oil components were 6-methyl-5-hepten-2-one (42.9%), β -caryophyllene (7.7%), bicyclogermacrene (5.1%), δ -cadinene (4.9%), and (*E*)-nerolidol (4.8%).

Keywords: Lauraceae, host plant, 6-methyl-5-hepten-2-one, sesquiterpenes

1. Introduction

Lindera benzoin (L.) Blume var. *pubescens* (Palmer & Steyermark) Rehd. "northern spicebush", is a shrub, up to 5 m tall, dioecious. Twigs are olive-green to brown with numerous light lenticels; leaves are alternate, simple, entire, glabrous or pubescent, elliptic to obovate, $7-13 \times 2-6$ cm, emit a spicy odor when crushed; the bark is brown to gray-brown with light colored lenticels; flowers, in axillary clusters, are small, pale green to yellow; fruit is a bright red oblong drupe, about 10 mm long ^[1, 2]. *L. benzoin* ranges in moist forests of eastern North America from southern Ontario to Florida. The plant has been used in Cherokee traditional medicine as a tea for treating coughs and colds ^[3] and is a host plant for the spicebush swallowtail butterfly, *Papilio troilus* L ^[4]. The essential oil compositions from leaves, twigs, and fruits from cultivated shrubs from Delaware have been reported ^[5]. This report presents the leaf essential oil composition from an individual growing in Huntsville, Alabama.

2. Materials and Methods

2.1 Plant Material

Leaves of *L. benzoin* were collected from a male individual shrub growing in Huntsville, Alabama (34° 38' 46'' N, 86° 33' 27'' W, 187 m elevation) on May 24, 2006. The plant was identified by W.N. Setzer, and a voucher specimen (LIBE-2006-WNS) has been deposited in the University of Alabama in Huntsville Herbarium. The fresh leaves were chopped and hydrodistilled for 4 h using a Likens-Nickerson hydrodistillation apparatus with continuous extraction with CHCl₃ (50 mL). The chloroform was evaporated to give the leaf essential oil: 48.33 g fresh leaves gave 74.8 mg (0.155% yield) essential oil.

2.2 Gas Chromatographic - Mass Spectral Analysis

The leaf essential oil of *L. benzoin* was subjected to gas chromatographic-mass spectral analysis using an Agilent 6890 GC with Agilent 5973 mass selective detector, fused silica capillary column (HP-5ms, 30 m × 0.25 mm), helium carrier gas, 1 mL/min flow rate; injection temperature 200 °C, oven temperature program: 40 °C initial temperature, hold for 10 min; increased at 3 °C/min to 200 °C; increased 2°/min to 220 °C, and interface temp 280 °C; EIMS, electron energy, 70 eV. The sample was dissolved in CHCl₃ to give a 1% w/v solution; 1-µL injection using a splitless injection technique was used. Identification of oil components was achieved based on their retention indices (determined with reference to a homologous series of normal alkanes), and by comparison of their mass spectral fragmentation patterns with those reported in the literature ^[6] and stored on the MS library [NIST database (G1036A, revision D.01.00)/Chemstation data system (G1701CA, version C.00.01.08)].

Correspondence: William N Setzer Department of Chemistry, University of Alabama in Huntsville Huntsville, AL 35899, USA wsetzer@chemistry.uah.edu

3. Results and Discussion

The chemical composition of *Lindera benzoin* leaf essential oil is presented in Table 1. A total of 37 compounds were identified in the leaf oil accounting for 94.6% of the total composition. *L. benzoin* leaf oil as revealed in this study was dominated by 6-methyl-5-hepten-2-one (42.9%), β -caryophyllene (7.7%), bicyclogermacrene (5.1%), δ -cadinene (4.9%), (*E*)-nerolidol (4.6%), (*E*)- β -farnesene (3.5%), and an unidentified sesquiterpene alcohol (3.8%). The chemical composition is qualitatively similar to the leaf oils of *L. benzoin* var. *benzoin* from Delaware and from Oregon ^[5], but with notable quantitative differences. 6-Methyl-5-hepten-2-one was

particularly abundant in the sample from Delaware (34.8%), as was β -caryophyllene (48.4%) and (*E*)-nerolidol (12.1%). The leaf oil from Oregon, however, had a much lower concentration of 6-methyl-5-hepten-2-one (1.9%). The different compositions can be attributed to very different geographical locations and climatic conditions. *L. benzoin* is a dioecious shrub. The samples from Delaware were female (fruits were collected) ^[5], but the gender of the shrub from Oregon was not reported ^[5]. The shrub from Alabama, this work, is male, and gender differences may also contribute to the qualitative differences in essential oil composition.

RI ^a	Compound	%	RI	Compound	%
860	cis-3-Hexenol	0.90	1504	cis-a-Bisabolene	0.20
979	Sabinene	0.47	1511	β-Bisabolene	2.84
995	6-Methyl-5-hepten-2-one	42.94	1514	γ-Cadinene	1.64
1011	α-Phellandrene	1.81	1525	δ-Cadinene	4.92
1028	<i>p</i> -Cymene	0.25	1537	α-Cadinene	0.15
1034	1,8-Cineole	1.18	1543	trans-a-Bisabolene	0.48
1106	6-Methyl-3,5-heptadien-2-one	0.29	1567	(E)-Nerolidol	4.57
1155	Citronellal	0.53	1576	Unidentified ^b	3.75
1231	Citronellol	0.48	1581	Caryophyllene oxide	0.98
1336	δ-Elemene	0.28	1641	τ-Cadinol	1.17
1356	Citronellyl acetate	0.22	1645	α-Muurolol	0.21
1384	β-Bourbonene	0.16	1654	α-Cadinol	1.22
1392	β-Elemene	0.98	1684	α-Bisabolol	1.32
1409	α-Gurjunene	0.17	1722	(E,E)-Farnesol	1.87
1420	β-Caryophyllene	7.74	1784	β-Bisabolenol	2.33
1436	α-trans-Bergamotene	1.14	1816	(Z,E)-Farnesyl acetate	0.85
1452	α-Humulene	0.70		Monoterpene hydrocarbons	2.53
1460	(E) - β -Farnesene	3.48		Oxygenated monoterpenoids	2.40
1476	γ-Muurolene	0.25		Sesquiterpene hydrocarbons	30.99
1480	Germacrene D	0.13		Oxygenated sesquiterpenoids	18.27
1497	Bicyclogermacrene	5.12		Others	44.13
1500	α-Muurolene	0.62		Total Identified	94.57

Table 1: Chemical composition of *Lindera benzoin* leaf essential oil.

^a RI = Retention Index, determined with reference to a homologous series of normal alkanes on an HP-5ms column.

^b Unidentified sesquiterpene alcohol, MS (EI): 222(0.3%), 204(26%), 189(7%), 161(100%), 147(6%), 133(18%),

119(39%), 105(61%), 91(38%), 81(31%), 67(9%), 55(12%).

It is not currently known if volatile leaf compounds serve to attract P. troilus adults to their host plants. It is known that tarsal contact of Lepidopterans with the host plant surface is necessary to initiate oviposition [7]; contact chemoreceptors on the tarsi are stimulated by particular phytochemicals and elicit oviposition [8-10], but host plant volatiles as well as visual cues likely contribute to host plant selection in Papilio spp.^[11, 12]. Host plants of P. troilus are restricted to the Lauraceae and include Sassafras albidum, Lindera melissifolia, and Persea borbonia in addition to L. benzoin [13]. S. albidum leaf oils are dominated by geranial (11-27%) and neral (10-18%), and β carvophyllene (5-13%), with lesser amounts of (E)-nerolidol and δ -cadinene, but no detectable quantities of 6-methyl-5hepten-2-one or bicyclogermacrene [14]. P. borbonia leaf oil, on the other hand, is dominated by camphor (35%) and 1,8-cineole (18%) ^[15]. There have apparently been no reports on leaf essential oil composition of L. melissifolia, but the volatiles from the fruits have been reported ^[16].

4. Conclusions

The leaf essential oil composition of *Lindera benzoin* var. *pubescens* from north Alabama has been determined. The oil was dominated by 6-methyl-5-hepten-2-one (44%), with lesser quantities of β -caryophyllene (8%), bicyclogermacrene (5%), δ -cadinol (5%), and (*E*)-nerolidol (5%). This work complements a

previous study of *L. benzoin* var. *benzoin* from Delaware and Oregon ^[5].

5. Acknowledgments

I am grateful to an anonymous private donor for the gift of the GC-MS instrumentation.

6. References

- 1. Brown CL, Kirkman LK. Trees of Georgia and Adjacent States. Timber Press, Portland, OR, USA, 1990.
- 2. Foote LE, Jones SB. Native Shrubs and Woody Vines of the Southeast. Timber Press, Portland, OR, USA, 1989.
- Hamel PB, Chiltoskey MU. Cherokee Plants and Their Uses – A 400 Year History. Stephens Press, Asheville, NC, USA, 1975.
- 4. Nitao JK, Ayres MP, Lederhouse RC, Scriber JM. Larval adaptation to lauraceous hosts: Geographic divergence in the spicebush swallowtail butterfly. Ecology. 1991; 72:1428-1435.
- Tucker AO, Maciarello MJ, Burbage PW, Sturtz G. Spicebush [*Lindera benzoin* (L.) Blume var. *benzoin*, Lauraceae]: A tea, spice, and medicine. Economic Botany. 1994; 48:333-336.
- 6. Adams RP. Identification of Essential Oil Components by Gas Chromatography / Mass Spectrometry, 4th Ed. Allured

Publishing, Carol Stream, IL, USA, 2007.

- Renwick JAA, Chew FS. Oviposition behavior in Lepidoptera. Annual Review of Entomology, 1994; 39:377-400.
- Simmonds MSJ. Importance of flavonoids in insect-plant interactions: feeding and oviposition. Phytochemistry. 2001; 56:245-252.
- Ono H, Yoshikawa H. Identification of amine receptors from a swallowtail butterfly, *Papilio xuthus* L. cloning and mRNA localization in foreleg chemosensory organ for recognition of host plants. Insect Biochemistry and Molecular Biology. 2004; 34:1247-1256.
- Heinz CA, Feeny P. Effects of contact chemistry and host plant experience in the oviposition behaviour of the eastern black swallowtail butterfly. Animal Behaviour. 2005; 69:107-115.
- Saxena KN, Goyal S. Host-plant relations of the citrus butterfly *Papilio demoleus* L. Orientational and ovipositional responses. Entomologia Experimentalis et Applicata. 1978; 24:1-10.
- 12. Mechaber WL, Capaldo CT, Hildebrand JG. Behavioral responses of adult female tobacco hornworms, *Manduca sexta*, to hostplant volatiles change with age and mating status. Journal of Insect Science. 2002; 2:1-8.
- Lederhouse RC, Ayres MP, Nitao JK, Scriber JM. Differential use of lauraceous hosts by swallowtain butterflies, *Papilio troilus* and *P. palamedes* (Papilionidae). Oikos. 1992; 63:244-252.
- 14. Kaler KM, Setzer WN. Seasonal variation in the leaf essential oil composition of *Sassafras albidum*. Natural Product Communications. 2008; 3:829-832.
- Tucker AO, Maciarello MJ, Wofford BE, Dennis WM. Volatile leaf oils of *Persea borbonia* (L.) Spreng, P. humilis Nash, and *P. palustris* (Raf.) Sarg. (Lauraceae) of North America. Journal of Essential Oil Research. 1997; 9:209-211.
- Oh J, Bowling JJ, Carroll JF, Demirci B, Başer KHC, Leininger TD, Bernier UR, Hamann MT. Natural product studies of U.S. endangered plants: Volatile components of *Lindera melissifolia* (Lauraceae) repel mosquitoes and ticks. Phytochemistry. 2012; 80:28-36.